Applications for X-Ray Spectroscopy

Jordan Planillo

Rensselaer Polytechnic Institute

Dr. Jerry Seidler

University of Washington

Abstract

X-ray crystallography via X-ray diffraction has been a reliable method method for determining crystal structure and composition. For complex crystals, as in the case of metalloproteins, X-ray diffraction fails to provide such information. X-ray spectroscopy gives better insight to the structure and composition of such complex crystal. The metalloprotein of interest is nitrogenase, the enzyme that allows for nitrogen fixation in plants and bacteria. This report details the experimental procedures for analyzing nitrogenase, specifically: experimental apparatus, simulation of nitrogenase with Fe(VI)N, and data analysis.

Introduction

With respect to metalloproteins, one of the hardest technical issues is to identify the existence and species of light elements bonded to catalytically active metal sites. X-ray diffraction cannot be used to obtain such information due to practical and experimental limitations. In terms of practical limitations, large amounts of high quality crystallized material are required. Such large crystals place experimental limitations such as crystal defects and the inability to detect light elements due to the presence of heavier elements.

Nitrogenase is one such metalloprotein of scientific interest as it is the only know method for nitrogen fixation and the exact details as to how nitrogenase performs this function are still unknown. The understanding of this function will allow for the efficient production of man-made fertilizers. The following chemical reaction details nitrogen fixation:

$N_2 + 8$ **H**⁺ + 8 **e**^{$-$} + 16 ATP \rightarrow 2 NH₃ + H₂ + 16 ADP + 16 P_i

Although the reaction of $N_2 + H_2 \rightarrow NH_3$ is energetically favorable, the N_2 triple bond must first be broken. The Mo-Fe co-factor in nitrogenase is the active catalytic site which strips electrons one at a time from N2, thus allowing it to dissociate and react with H2 to form NH3. Einsle, et al, Science (2002) (Fig.1), proposes a new model for the structure of the catalyticallyactive Mo-Fe cofactor where there is a light atom (C, N, O, or S) in a central cavity of the metal cluster, it is suspected to be nitrogen as it can play the role of a charge reservoir.

X-ray spectroscopy is the method of choice for studying metalloproteins because the xray emission spectra from metal atoms allow for the identification of light ligands. The emission of interest is when semicore electrons of the ligand fill in the 1s hole in the metal- the Kβ'' emission. The Kβ'' emission energy is directly correlated to the chemical species of the light ligand, however it is very hard to detect (Fig. 2).

U. Bergmann, C.R. Horne, T.J. Collins, J.M. Workman, S.P. Cramer, Chem. Phys. Lett. 302 (1999) 119.

Fig .2

Experimental Setup

Before any measurements can be taken, calculations must first be made. One critical calculation is determining the correct crystal for the photon emission energy of interest. This is obtained using Bragg's law:

$$
2d\sin(\theta)=n\lambda
$$

From this, a table relating emission energies to analyzer crystals was compiled (Table 1). For the case of nitrogenase, the emission energy of Fe Kβ is 7057.980 eV with the corresponding crystal of Ge(620).

Another critical piece of experimental hardware that I was heavily involved in, was the sample box. Special design considerations included the ability for the sample box to contain a He gas environment and accommodate a homologated sample holder. The sample box was manufactured by milling aluminum square tubing.

Fig. 3: Sample box (Design and Construction)

Additional preparation (assembly/sample preparation/sealing/sheilding)took place on-site at the Advanced Photon Source at Argonne National Laboratory.

Fig.4: Spectrometer Assembly

Fig. 5:Noise Reduction

He gas environment in the spectrometer (left), in sample box (center), Lead shield (right)

Fig. 6: Raw Spectrometer data

Fe Elastic scattering lines (bottom) and fluorescence lines (top)

Fig. 7: Calibriation Matrix

Using the elastic scattering lines, the energies associated with each pixel can be assigned

Fig. 8: Nitrogenase Simulation with Fe(VI)N

The purpose of this measurement is to observe the Fe $K\beta$ " in a sample similar to nitrogenase and determine systematic considerations for nitrogenase in future experiments. The initial plot (blue line) is the initial 30s exposure.

Analysis

Fig. 8 shows that each subsequent exposure to the beam changes the $Fe(IV)N$ spectrum. This is most evident in the Fe $3p\rightarrow1s$ transition, where there is a large increase in Fe spin. This spin is likely explained by spin trapping (Vanko, et al, Angew. Chem. Int. Ed. 2007), where high energy excitations cause a cascade of ionization in the sample. As the cascades settle down, the valence electrons are captured in a high spin state.

Fig. 10: Spin trapping

Fig. 9 provides additional detail on the effects of beam exposure on the sample, specifically, on the presence of the Kβ'' emission. The 10 minute exposure clearly shows the Kβ'', but not so much on the shorter exposures. The dramatic change in spectra indicates that beam damage is evident and should be taken into consideration during future experiments.

Conclusions

The study of nitrogenase has been greatly aided by experimenting on. Fe(VI)N. Most important is the indication of beam damage on the Fe(IV)N. Additionally, it was possible to observe the faint emission of $K\beta$ ", giving further insight as to what might be contained in nitrogenase's MoFe cofactor. For future experiments, one must take beam damage countermeasures. Such countermeasures could include cooling the sample to counter act small damages and possibly sweeping the beam over the sample to prevent large scale damage from prolonged exposure.

Table 1: Emission line energy and corresponding crystals

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